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### 19 NARCOTIC METHODOLOGY

- **19.1 Brief Pharmacology:** Analgesic, sedative effects and causes constipation.
- **19.2 Drug Group Examples:** Morphine, heroin, hydromorphone, pentazocine, codeine, hydrocodone, oxycodone, methadone, propoxyphene, pethidine (meperidine), and fentanyl

# 19.3 Scheduling:

- Schedule I heroin
- Schedule II morphine, oxycodone, hydromorphone, methadone, pethidine (meperidine), fentanyl, codeine (pure), hydrocodone (pure), levomethorphan, 6-monoacetylmorphine (morphine derivative)
- Schedule III Some preparations of hydrocodone or codeine
- Schedule IV pentazocine
- Schedule V Some preparations of codeine (usually cough preparations)
- Non-controlled dextromethorphan
- Appropriate caution must be exercised when determining the control status of compounds listed in multiple schedules.
   Any questions should be answered by consulting appropriate compendia references such as the PDR, Poison Control,
   DEA Logo Index and the appropriate state or federal codes. Marked capsules or tablets need not be quantitated.
   Questionable samples require at least a semi-quantitative workup to determine the schedule.

## 19.4 Extraction:

- 19.4.1 Most narcotics may be extracted from basic solutions with organic solvents.
- 19.4.2 Morphine may be extracted from aqueous solution by the addition of a carbonate/bicarbonate buffer and extracting with CHCl<sub>3</sub> or CHCl<sub>3</sub>/isopropanol (8:2). Morphine sulfate is not soluble in chloroform.
- 19.4.3 Methadone is often found in orange juice or Tang-type orange drink. The solution should be made basic with sodium carbonate and extracted into CHCl<sub>3</sub> or CH<sub>2</sub>Cl<sub>2</sub>.
- 19.4.4 Narcotics in cough syrups may be extracted from basic solutions with organic solvents.
- **19.5 Color Test Results:** Most narcotics give colors with Marquis, Meckes and Froehdes reagents (see Table 1). Numerous other materials give similar colors, such as methapyriline, glycerol guaiacolate (guaifenesin), and pyrilamine, and are also included.

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TABLE 1: Positive Color Test Reactions

Compound	Marquis	Meckes	Froehdes	HNO <sub>3</sub>
Morphine	Purple	Green	Purple	Red
Heroin	Purple	Green	Purple	Yellow
Codeine	Purple	Blue-green	Green	Yellow
Propoxyphene	Black	Orange/brown	Brown	No reaction
Meperidine	Orange	Yellow-green	Grey	
Pentazocine	Red→olive green	Olive green	Blue	Yellow
Hydromorphone	Yellow→red	Yellow-orange	Blue→purple	Yellow-orange
Hydrocodone	Purple	Green	Lt. yellow	No reaction
Oxycodone	Yellow <b>→</b> purple	Yellow→olive	Yellow	No reaction
Methadone	Slow pink	Yellow- green→green		
Methapyriline	Purple	Purple	Purple	No reaction
Guaifenesin	Reddish purple	Green/purple	Green with purple streaks	Yellow
Pyrilamine	Purple	Purple	Purple	

### 19.6 TLC:

- 19.6.1 Baths: TLC1, TLC2, TLC3, TLC4 and TLC5 are recommended.
- 19.6.2 Detection sprays:
  - 19.6.2.1 Iodoplatinate
  - 19.6.2.2 Ceric Sulfate Fentanyl may require the use of ceric sulfate as an overspray due to the minute amounts of this material found in most pharmaceutical preparations.

### 19.7 UV:

- 19.7.1 Not usually unique to the specific compound.
- 19.7.2 Results:
  - Morphine max. at 285 nm in acid
  - Heroin max. at 280 nm in acid (Heroin will often hydrolyze to monoacetylmorphine and morphine with prolonged exposure to aqueous solutions.)
  - Methadone max. at 250 and 292 nm in acid
  - Pethidine (Meperidine) triplet max. at 258 nm in acid

# 19.8 Dextromethorphan Enantiomer Determination

- 19.8.1 Because of the scheduling differences between levomethorphan (Schedule II) and dextromethorphan (non-controlled), a microcrystal test must be performed for differentiation unless found in a recognizable pharmaceutical preparation.
- 19.8.2 Materials:
  - Glass slides and coverslips
  - Polarizing microscope
  - 10% Platinic Chloride solution in water (w/v)
  - 1% acetic acid solution in water

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• Dextromethorphan reference standard

### 19.8.3 Procedure:

- 19.8.3.1 Place one drop of 1% acetic acid in water to a small portion (less than 1mg of pure dextromethorphan is needed) of the dextromethorphan standard on a glass slide. Then add a portion of a drop of 10% platinum chloride in water and cover. Feathery dendrites will form within 2 minutes.
- 19.8.3.2 Place one drop of 1% acetic acid in water to a small portion of the sample on a glass slide. Then add a portion of a drop of 10% platinum chloride in water and cover. Feathery dendrites will form when the sample is a pure enantiomer. A racemic mixture will not form these microcrystals.
- 19.8.3.3 To verify the identity as the dextro isomer, mix a small amount of sample with approximately the same amount of dextromethorphan reference standard on a glass slide. Add the acetic acid and platinum chloride solutions as stated above. Observe any microcrystalline formation. If the sample is dextromethorphan, the feathery dendrites will form; if the sample is levomethorphan or racemethorphan, no dendrite-shaped crystals will form.
- 19.8.3.4 The microcrystal formations should be contemporaneously verified or photographed. Verification must be documented in the case notes with the initials of the verifier and the date.
- 19.8.3.5 The sample may need to be purified to allow for crystal formation. Mixtures of dextromethorphan and MDMA will need to be separated before the crystal test, as pure MDMA reacts with platinum chloride to form similar, but not the same, microcrystals.
- 19.8.3.6 Reference: Fulton, Charles C. *Modern Microcrystal Tests for Drugs*, New York: Wiley-Interscience, 1969, pp. 58-59.

### 19.9 Codeine Quantitation

19.9.1 See GC section 10 for general quantitation procedure.

### 19.9.2 Reagents:

- Methylene Chloride or Chloroform
- Dicyclohexylphthalate (DCHP)
- Codeine Standard

### 19.9.3 Internal Standard Solution:

- 19.9.3.1 Prepare a sufficient volume to dilute the codeine standard solution and all samples.
- 19.9.3.2 Prepare a 1.5 mg/mL solution of DCHP in methylene chloride or chloroform in an appropriate volumetric flask.

### 19.9.4 Codeine Standard Solutions:

- 19.9.4.1 Weigh 20 mg of Codeine and transfer to a 10 mL volumetric flask with internal standard solution. Dilute to mark with internal standard solution. This results in a solution for 2.0 mg/mL codeine in internal standard solution.
- 19.9.4.2 Prepare a solution of another concentration within the linear range in the same manner to use as the check standard.

# 19.9.5 Sample preparation:

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- 19.9.5.1 Verify the identity of the salt originally present in the sample by suitable chemical testing. Adjust sample weight accordingly.
- 19.9.5.2 Weigh approximately 20 mg of the sample and made basic with 4N sodium hydroxide solution until pH is above 10.
- 19.9.5.3 Extract with a known volume of internal standard solution in a small separatory funnel.
- 19.9.5.4 Extraction is necessary in order to remove the salt for comparison to the Codeine standard solution.

# 19.9.6 GC parameters:

• Column: 15 m HP-1 or HP-5 capillary (0.25 mm i.d., 0.25 µm film thickness)

Oven temperature: 220-240° C
FID temperature: 280° C

19.9.7 Codeine elutes prior to DCHP.

### 19.10 Heroin Quantitation

19.10.1 See GC section 10 for general quantitation procedure.

### 19.10.2 Reagents:

- Methylene Chloride or Chloroform
- Dicyclohexylphthalate (DCHP).
- Heroin Standard

### 19.10.3 Internal Standard Solution:

- 19.10.3.1 Prepare a sufficient volume to dilute the heroin standard solution and all samples.
- 19.10.3.2 Prepare a 1.0 mg/mL solution of DCHP in methylene chloride or chloroform in an appropriate volumetric flask.

### 19.10.4 Heroin Standard Solutions:

- 19.10.4.1 Weigh ~20 mg of Heroin and transfer to a 10 mL volumetric flask with internal standard solution. Dilute to mark with internal standard solution. This results in a solution of ~2.0 mg/mL Heroin in internal standard solution.
- 19.10.4.2 Prepare a solution of another concentration within the linear range in the same manner to use as the check standard.

# 19.10.5 Sample Preparation:

- 19.10.5.1 Weigh 10-40 mg of sample into a test tube.
- 19.10.5.2 Dilute with internal standard solution to appropriate volume.

### 19.10.6 GC parameters:

- Column: 15 m HP-1 or HP-5 capillary (0.25 mm i.d., 0.25 μm film thickness)
- Oven temperature: 260-280° C

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• FID temperature: 280° C		
19.10.7 DCHP elutes prior to Heroin. Peaks between DCHP and Heroin are off acetylcodeine.	ten due to monoacetylmorphine or	
	♦ Er	